



## Baird Parker Agar Base

**RDM-BPA-01**

### Principle

Baird Parker Agar was developed by Baird Parker (1962), composed of tryptone, meat extract (equivalent to beef extract), glycine, yeast extract, sodium pyruvate, lithium chloride and agar. Tryptone, meat extract and yeast extract provide nitrogen, carbon, sulphur and vitamins. Glycine and sodium pyruvate protect injured cells and helps to recovery them and stimulates the growth of *Staphylococcus aureus*. Lithium chloride and potassium tellurite acts as inhibitor agent for contaminating microflora. The tellurite additive is toxic to egg yolk-clearing strains other than *S. aureus* and imparts a black color to the colonies.

**Use:** Recommended for the isolation, enrichment and enumeration of coagulase positive staphylococci from food and clinical samples.

### Contents\*

<b>Ingredients</b>	<b>Gram/Litre</b>
Tryptone	10.000
Meat Extract	5.000
Yeast Extract	1.000
Glycine	12.000
Sodium Pyruvate	10.000
Lithium Chloride	5.000
Agar	20.000
pH at 25°C	6.9 ±0.2

\* Formula adjusted for optimum performance and parameters

# Equivalent to beef extract

**Directions:** Dissolve 63.00 grams in 950 ml distilled water, boil to dissolve the medium completely and sterilize by autoclaving at 15 lbs pressure (121 °C) for 15 min, cool it to 42-45 °C. After cooling to 45-50°C, add 50 mL of Egg Yolk Tellurite Supplement and 3 ml sterile 3.5% Potassium Tellurite solution or 50 ml Egg Yolk Tellurite Emulsion. Mix thoroughly before dispensing.

### Specimens types analyzed

Pharmaceutical samples, clinical and non-clinical samples etc.

### Precautions to be taken

These microbial media are intended for the in-vitro use only. All the handling, experiments, storage, and discarding should be performed with the help of skilled and knowledgeable technicians and as per the established guidelines. The material should be disposed only after proper sterilization by autoclaving. Please go through the MSDS of the media to avoid any accidents or in emergency.

### Performance and Evaluation

The expected performance of the medium is liable to use as per the direction on the label when stored at optimum conditions and within expiry date.

### Quality Control

<b>Appearance</b>	Beige colored free flowing, homogeneous powder
<b>Reaction of 6.3% solution</b>	6.9 ±0.2 at 25 °C
<b>pH</b>	6.60- 7.00
<b>Gelling</b>	Firm comparable with 2% agar gel

<b>Color and clarity of ready medium</b>	Light yellowish colored opalescent gel
<b>Growth Promotion properties</b>	Best at $\leq 100$ CFU at 32-37 °C for 18-72 h
<b>Indicative properties</b>	Optimum at $\leq 100$ CFU at 32-37 °C for 18-48 h
<b>Negative control</b>	Performed using sterile distilled water

### Different Microbial Response

Organism	Inoculum	Growth	Recovery	Colony Color	Lecithinase
<i>Staphylococcus aureus</i> (ATCC 6538)	50-100	Luxurious	60-70%	Gray black shiny	Positive opaque zone observed
<i>Proteus mirabilis</i> (ATCC25933)	50-100	Luxurious	70-80%	Brown black	Negative
<i>Escherichia coli</i> (ATCC 8739)	50-100	Poor	0-10%	Brown black	Negative

### Storage and Shelf Life

Hygroscopic; keep container tightly closed. Store in cool dry place.

**Disposal:** To avoid the contamination or propagation of any hazardous microbes the used, unusable or modified preparation of this product must be disposed after autoclaving after completion of task.

### Reference

1. Atlas, R. M. (2005). *Handbook of media for environmental microbiology*. CRC press.
2. *Difco Manual* (1998). 11th Edition. Difco Laboratories., Division of Becton Dickinson and Company, Sparks, Maryland, USA.
3. Rand, M. C., Arnold E. Greenberg, and Michael J. Taras. (1976) *Standard methods for the examination of water and wastewater*. Prepared and published jointly by American Public Health Association, American Water Works Association, and Water Pollution Control Federation., 1976.
4. Salfinger Y., and Tortorello M.L., (2015), *Compendium of Methods for the Microbiological Examination of Foods*, 5<sup>th</sup> Ed., American Public Health Association, Washington, D.C.
5. Wehr H. M. and Frank J. H., (2004), *Standard Methods for the Microbiological Examination of Dairy Products*, 17<sup>th</sup> Ed., APHA Inc., Washington, D.C.

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