



## Chromogenic UTI Agar

**RDM-UTI-01**

### Principle

Chromogenic UTI agar is used for presumptive identification of microorganisms causing urinary tract infections. The organisms mainly responsible for urinary tract infections are *Escherichia coli*, *Staphylococcus saprophyticus*, *Klebsiella species*, *Pseudomonas aeruginosa*, *Enterococcus faecalis*, *Proteus mirabilis*, and other coliforms. Media is composed of peptone, chromogenic mixture and agar. Peptone provides nitrogenous and long chain amino acids. The chromogenic mixture contains chromogens and nutrients, the nutrients provide nitrogenous and carbonaceous compounds, vitamins and essential nutrients including tryptophan. While the chromogenic substance used in this media are X-Glucoside, Red-β-D-galactopyranoside and isopropylthio-β-galactoside. X-Glucoside is a substrate for β-Glucosidase that, upon enzymatic action, gives an insoluble indigo-blue chromophore. Red-β-D-galactopyranoside is used for detection of beta-galactosidase. IPTG (isopropylthio-β-galactoside) is an inducer of β-galactosidase activity in bacteria and is suitable for use with X-gal or Red-gal to detect lac gene activity in *E. coli* or genetically modified microorganisms. The medium also contains tryptophan which acts as an indicator of tryptophan deaminase activity.

The *Escherichia coli*, produce β-galactosidase enzyme activity, cleave red-β-D-galactopyranoside and forms pink color colonies. The enterococci cleave X-glucoside, by producing β-glucosidase enzyme, and result in formation of blue colonies. However, some members of the coliform group, cleaves both the chromogen to form purple colonies. While *Proteus*, *Morganella* and *Providencia* spp. are differentiated on basis of tryptophan deaminase activity and generally forms brown color colonies. Some *Enterobacter cloacae* lack β-glucosidase, resulting in pink colonies, similar to *Escherichia coli*, and further differentiated on basis of indole production by using Kovac's reagent.

**Use:** Recommended for presumptive identification of microorganisms causing urinary tract infections.

### Contents\*

| Ingredients         | Gram/Liter |
|---------------------|------------|
| Peptone             | 15.000     |
| Tryptophan          | 2.000      |
| Chromogenic mixture | 15.500     |
| Agar                | 15.000     |
| pH at 25°C          | 6.8 ±0.2   |

\* Formula adjusted for optimum performance and parameters

**Directions:** Dissolve 47.50 grams in 1000 ml distilled water. Boil to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Cool to 45-50°C. Mix well and pour into sterile petri plates.

### Specimens types analyzed

Clinical samples: urine, faeces, Water samples and Food samples.

### Precautions to be taken

All the handling, experiments, storage, and discarding should be performed with the help of skilled and knowledgeable technicians and as per the established guidelines. The material should be disposed only after proper sterilization by autoclaving. Please go through the MSDS of the media to avoid any accidents or in emergency.

## Performance and Evaluation

The expected performance of the medium is liable to use as per the direction on the label when stored at optimum conditions and within expiry date.

## Quality Control

|                                   |                                                |
|-----------------------------------|------------------------------------------------|
| Appearance                        | Light beige colored, free-flowing, homogeneous |
| Reaction of 4.75 % solution       | 6.8 ±0.2 at 25 °C                              |
| pH                                | 6.60- 7.00                                     |
| Gelling                           | Firm comparable with 1.5% agar gel             |
| Color and clarity of ready medium | Light beige, clear opalescent gel              |
| Growth Promotion properties       | Best at ≤ 100 CFU at 32-37 °C for 18-72 h      |
| Indicative properties             | Optimum at ≤ 100 CFU at 32-37 °C for 18-48 h   |
| Negative control                  | Performed using sterile distilled water        |

**Different Microbial Response:** Cultural characteristics observed after incubation at 33-37°C for 18-24 hours. Inoculum 50-100 CFU.

| Organism                      | ATCC  | Growth    | Recovery | Colony color                  |
|-------------------------------|-------|-----------|----------|-------------------------------|
| <i>Escherichia coli</i>       | 25922 | Luxuriant | ≥ 60%    | Pinkish purple                |
| <i>Pseudomonas aeruginosa</i> | 27853 | Luxuriant | ≥ 60%    | Colorless or greenish pigment |
| <i>Salmonella typhimurium</i> | 14028 | Luxuriant | ≥ 60%    | Colorless                     |
| <i>Staphylococcus aureus</i>  | 25923 | Luxuriant | ≥ 60%    | Normal pigmentation           |
| <i>Klebsiella aerogenes</i>   | 13048 | Luxuriant | ≥ 60%    | Purple                        |
| <i>Proteus mirabilis</i>      | 12453 | Luxuriant | ≥ 60%    | Cream color with brown halo   |
| <i>Enterococcus faecalis</i>  | 14506 | Luxuriant | ≥ 60%    | Blue                          |

**Storage and Shelf Life:** The product is highly hygroscopic; keep the container tightly closed at all times and store it properly as per the conditions mentioned on the label. The declared expiry is valid only when stored as per the conditions mentioned on the label.

**Note:** Sterilize media immediately after reconstitution.

**Disposal:** To avoid the contamination or propagation of any hazardous microbes the used, unusable or modified preparation of this product must be disposed after autoclaving after completion of task.

## Reference

1. Baird R.B., Eaton A.D., and Rice E.W., (Eds.), 2015, Standard Methods for the Examination of Water and Wastewater, 23rd ed., APHA, Washington, D.C.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.
3. Jorgensen, J. H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015), Manual of Clinical Microbiology, 11<sup>th</sup> Edition. Vol. 1 Wastewater, 23<sup>rd</sup> Ed., APHA, Washington, D.C.
4. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.

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